Review Article Physiology, environmental change, and anuran conservation

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Abstract

Physiology, environmental change, and anuran conservation. This paper reviews diverse ways in which the emerging discipline of conservation physiology may contribute to the conservation of anuran amphibians. We first present a summary of the goals of conservation physiology. Then, we review five specific sources of environmental change that are known to affect anurans: introduced pathogens (highlighting the case of chytridiomycosis) and exotic species, pollutants, habitat fragmentation, and global climate change. We discuss these types of environmental change in the context of how and why altered environments may become stressing to anurans and cause population declines. We also discuss synergism among variables that may worsen the effect of environmental alterations, and emphasize the importance of conservation physiology for Brazilian anurans.

Keywords: Anura, conservation physiology, environmental change, chytridiomycosis, exotic species, pollutants, habitat fragmentation, global climate change.

1. Introduction

Comparative physiology has a long-standing history of collateral contributions to conservation biology, but recently the specific field of conservation physiology has grown to be considered an independent emerging discipline (Wikelski and Cook 2006). An important goal of this new field is to understand the ability of organisms to deal with environmental change, including those resulting from human influence.

Received 14 June 2007. Accepted 21 November 2007. Distributed December 2007. Although human-induced changes tend to be very rapid and have the potential to affect all organisms, some taxonomic groups are far more susceptible to environmental change than others. Anuran amphibians figure among the most vulnerable taxa, as deduced from current dramatic rates of population and species extinction. The decline of anuran populations and its relationships with ecological change have been the focus of divergence regarding proper actions to be taken (Mendelson et al. 2006, Parmesan 2006, Pounds et al. 2006a), a debate to which conservation physiology may contribute significantly. In this paper we discuss how physiological studies may bolster the cause of anuran conservation and management.

2. A synopsis on the goals and principles of conservation physiology

The possible outcomes of noxious environmental change to organisms are local extinction, migration, or adjustment to the new condition (Donnelly 1998, Helmuth et al. 2005). Adjustment, in this case, means a modification of behavior or physiology that ameliorates the effect of a stressful change in environmental conditions. Physiological adjustments are often the only mechanism by which organisms facing environmental change can maintain homeostasis, the state of internal equilibrium required for life (Carey 2005, Wikelski and Cooke 2006). The ability to maintain homeostasis through time is intuitively related to the life-time ecological success of organisms exposed to environmental change (McEwen and Wingfield 2003), and is called allostatic capacity (Wikelski and Cooke 2006). The point at which the threshold for stress tolerance of a population is surpassed is known as allostatic overload. This is a valuable concept in conservation physiology because it relates stress to a physiological state more than to the type, magnitude, or time-scale of the stressor (Wikelski and Cooke 2006).

Differences in allostatic capacity result from physiological adjustments occurring at various levels of organization (e.g., individuals, populations, and species) and time scales (Wikelski and Cooke 2006). Within the lifetime of an individual, for example, fully reversible adjustments may occur through the process of acclimatization. Likewise, modifications of physiology, reversible or not, may occur through development and under the influence of the environment, and are associated with the concept of phenotypic plasticity. Finally, adjustments leading to shifts in allostatic capacity may take place across generations in the context of evolutionary processes. Many examples of the importance of these processes exist in the literature. Acclimatization to temperature, for example, seems fundamental

for crabs to adjust to global warming (Stillman 2003), and directional changes in the frequency of alleles linked to stress physiology, and attributable to climate change, have been observed in mountain insects (Rank and Dahlhoff 2002). The physiological tolerance of insects to humidity and temperature is related to geographic range, and apparently shifts in association with climatic change (Karban and Strauss 2004).

Three goals of conservation physiology that we believe are of particular importance are 1) to understand the scope and paths through which organisms facing noxious environmental change adjust their physiology to overcome stress (Wikelski and Cooke 2006); 2) to reveal the physiological challenges that affect critical parts of the life cycles of organisms (Helmuth et al. 2005); and 3) to evaluate the costs and tradeoffs associated with physiological adjustments. These costs may be direct, in the sense that noxious environmental change may either increase the energy requirements of animals or decrease their ability to obtain energy (Carey 2005, Wikelski and Cooke 2006). The cost of maintenance of amphibians, for example, may increase directly as a consequence of temperature shifts (see Section 3.2) or exposure to pollutants (see Section 3.4). Energy expenses may also increase indirectly, for example when environmental change results in increased need for locomotion (Carey 2005, Wikelski and Cooke 2006) (see Section 3.5). Conversely, pathological conditions (Parris et al. 2006) or experimental exposition to predators (Skelly 1992) may reduce the inclination for activity in anuran tadpoles. When environmental change downgrades energetic budgets, organisms need to reduce allocation to growth, storage, reproduction or activity (Calow 1991, Brauner et al. 1994, Hopkins et al. 1997, Rowe et al. 1998, Beyers et al. 1999, Barbieri et al. 2002). Energy, however, is not the only possible price of physiological adjustments. Additional potential costs include phenotypic trade-offs, that is, additional changes that occur in parallel with physiological adjustments, and that result from phenotypic correlations among traits (Angilletta *et al.* 2003, Helmuth *et al.* 2005). Phenotypic correlations may be based in genetic correlations. For example, selection favoring a reduction in the thermal dependency of developmental rate of *Rana sylvatica* results in a concomitant, and apparently unavoidable, thermal dependency of body size (Berven 1982).

Several fields of physiology have made important contributions to conservation, and interested readers should consult the reviews by Carey (2005), Helmuth et al. (2005), and Wikelski and Cooke (2006). In summary, endocrinology, ecological physiology, immunology, physiological genomics and other physiological disciplines have provided tools to evaluate stress, illustrate the control and triggers of reproduction, understand the patterns of animal distribution, and to comprehend the effects of physical variables on physiology. Evolutionary physiology has contributed greatly to understanding the factors that modulate and restrict the evolution of physiological traits (Angilletta et al. 2002), with concrete links with life-history traits (Sears and Angilletta 2004). Comparative and ecological physiology have helped in the understanding of acclimation, reversible physiological responses by which organism may adjust to environmental change such as global warming (Stillman 2003). The applications of physiology to anuran conservation are clearly many, and it would be beyond the scope of this paper to treat them all. Instead, we highlight in the next sections some scenarios that may interest a broad audience of herpetologists, and that illustrate how physiological studies may contribute to the conservation and management of endangered anuran amphibians.

3. Anurans and environmental change

In this section we focus on five categories of environmental degradation that are particularly important for anuran conservation. First, we focus on human-spread pathogens, specifically chytridiomycosis, the putative causal agent of the decline and extinction of several anuran populations and species (Daszak et al. 2001). The second category is global climate change, and the need to understand how anuran physiology would react to changes in mean temperature and thermal variance, and to extreme climatic events. Then, we describe briefly the importance of understanding synergistic effects among environmental variables, focusing on temperature and chytridiomycosis. Subsequently, we move to pollutants, which may become stressors, particularly at early developmental stages. Water pollution may have deleterious effects on anuran egg hatching, perhaps acting synergistically with other factors (Karasov et al. 2005). Next, we review habitat fragmentation, and discuss why survival in a fragmented forested environment may be influenced by the ability of individuals to cross gaps. This factor has been recognized as an important attribute for the survival of anuran populations (Zimmerman and Bierregaard 1986, Dale et al. 1994) but has received little formal attention. Finally, we center on the physiological and behavioral ability of exotic species to outcompete local species, which is another point of interface between conservation biology and physiology.

3.1. Chytridiomycosis

Chytridiomycosis is a disease caused by a fungus, *Batrachochytrium dendrobatidis*, and is, for many scientists, the causal agent explaining local anuran extinction (Berger *et al.* 1998, Collins 2003, Daszak *et al.* 2003, Lips *et al.* 2003a,b, Retallick *et al.* 2004, Lips *et al.* 2005, Woodhams and Alford 2005, Pounds *et al.* 2006a). The decline of anuran populations from chytridiomycosis has become a top issue in conservation biology, one to which investigations in comparative and ecological physiology have much to offer. Anuran species differ in their susceptibility to chytridiomycosis (Berger et al. 1998, Lips et al. 2003a,b, Retallick et al. 2004, Woodhams and Alford 2005, Woodhams et al. 2006a), and this variation apparently stems from differences in the immune system (Woodhams et al. 2006a). A main immune organ of anurans, generally the first one to enter in contact with B. dendrobatidis in the field, is the skin (Carey 2005). Therefore, a step to understand interspecific differences in susceptibility to chytridiomycosis is to perform comparative studies of the morpho-physiology of the anuran skin, and of the ecologically-relevant microbiological properties of its secretions. The anuran skin contains various types of glands, including the granular glands involved in the production of antimicrobial peptides. A large number of anuran-skin peptides and alkaloids (amines) have been characterized (see review in Pukala et al. 2006), but few species have been studied, particularly in Neotropical regions. These skin peptides, which vary in type and proportion among species, confer various degrees of protection from pathogens, including B. dendrobatidis (Rollins-Smith et al. 2002a,b, 2005, Rollins-Smith and Conlon 2005, Woodhams et al. 2006a) or bacteria isolated from natural habitats (Ashcroft et al. 2007). Further studies will help to understand the mechanisms underlying peptide action (Rollins-Smith et al. 2002a,b) and the eventual phylogenetic constraints related to chytridiomycosis vulnerability (Woodhams et al. 2006b).

Physiological ecology is relevant in the context of chytridiomycosis because a number of environmental factors that may further increase stress levels, including temperature, intensity of UV-B radiation and concentration of toxic chemicals, influence susceptibility to the disease (Johnson *et al.* 2003). Noxious environmental conditions may increase the chances of infection by depressing the protection derived from skin products (Rollins-Smith *et al.* 2002a,b), and by increasing the concentration of circulating glucocorticoids

(Simmaco *et al.* 1997, Matutte *et al.* 2000), stress-related steroid hormones derived from cholesterol that may be harmful when chronically elevated. Immunological depression may also have an indirect, but perhaps important, link with the biotic environmental. This is so because part of the chemicals produced by the anuran skin, alkaloids in particular, are sequestered from dietary sources including ants, beetles, millipedes, and other arthropods (Daly 1995). As a consequence, a depauperate ecological community may constrain the biochemical profile of anuran skin secretions.

3.2. Global climate change

The physiological adjustments required of the organisms in response to climate change is a cause of concern among scientists working on systematic groups as diverse as plants (Beardall et al. 1998), fish (D'Cruz et al. 1998), birds (Wolf 2000), and certainly, amphibians. Ever since the population crash of the Costa Rican golden toad, Ollotis periglenes, in 1987, attributed to erratic weather perhaps induced by global warming (Pounds and Crump 1994), herpetologists have been concerned about climate change and anuran conservation. From what we know from physiology, this concern is fully justified. Anurans experience pervasive effects of temperature on their behavioral and physiological function, including the immune system (Beecher and Demas 2004), and may respond to infection using behavioral thermoregulation. Not surprisingly, therefore, the intricate ways by which global climate change may affect anurans include interactions with other major problems such as chytridiomycosis (see next section). It is also essential to keep in mind that global climate change is not a smooth monotonic increase in environmental temperatures, but a complicated pattern in which emerging extreme events are of substantial importance (Marengo 2006, Mitchell et al. 2006). Droughts associated with global warming, for example, in combination with other factors such as fungal infections, may have been the cause of populational declines in Eleutherodactylus from Puerto Rico (Burrowes et al. 2004). Frosts possibly contributed to the decline of anuran communities in the Brazilian Atlantic Forest (Heyer et al. 1988; see Section 4), and a reduction of the amount of rain per year possibly caused the anuran population declines observed during long-term studies at the Savannah River Site (SRS) in South Carolina, USA (Daszak et al. 2005). It is impossible to prove that these past atypical climatic extremes actually caused the declines, and it would be even more difficult to prove that such extremes were a direct consequence of the global warming phenomenon. However, the data available do suggest that climate extremes may be consequential for anuran populations, and that the likelihood of such events to occur has increased with global warming. Understanding the scope and physiological basis for anuran tolerance to climate extremes seems an imperative issue in the agenda for anuran conservation.

An important concern when discussing the implications of climate change for anuran conservation regards their possibilities for behavioral and physiological adjustment to temperature change. Behavioral responses, such as a shift towards an earlier onset of calling activity during the day, do not compensate for reduced temperature in high-elevation Andean anurans (Navas 1996a). Some temperate anuran species appear to compensate for a warmer climate by shifting the time of year in which the reproductive season occurs (Beebee 1995), but no common trend exists, and a number of species in North-America and Europe have not adjusted reproduction in this manner (Blaustein et al. 2001, 2003, Beebee 2002, Corn 2003). In addition, even when present, behavioral adjustments may be insufficient to compensate for the effects of warming. Warmer winters, for example, affect the reproductive output of Bufo bufo by decreasing female body condition

during hibernation (Reading 2007). Warming may also affect the morphology of anurans, particularly body size. Males of the "Rana esculenta" complex (water frogs of Central Europe) exhibit body sizes that are up to 10% larger than those measured in 1963, a period over which indirect measures suggest that temperatures became more moderate. Females, in contrast, exhibited no or contradictory patterns, a finding that illustrates the complexity of temperature effects on anurans (Tryjanowski et al. 2006). Because temperature may affect the type and availability of prey items, which in turn relates to growth rates, temperature may also cause indirect effects on anuran adult size (Tryjanowski et al. 2006). Overall, physiological, more than behavioral adjustments appear to compensate for climate change in anurans.

Global climate change generates particular risk to species adapted to cool or to extreme environments. Some species from polar or high altitude environments, for example, already show drastic contractions of their natural ranges (Parmesan 2006). Altitude seems to be a factor increasing the risk of anuran population declines, although its possible effects are not simple in pattern (Pounds et al. 2006a). An example in which a concrete causative relationship between temperature and physiological stress has been proposed concerns montane salamanders from the Appalachian highlands (Desmognathus). These cold-adapted salamanders, perhaps due to evolutionary or physiological constraints, do not exhibit adjustments with respect to the effects of temperature on the cost of maintenance (metabolic rate) along an altitudinal gradient. Although the metabolic rate is temperature dependent between 5 and 15°C, as expected for amphibians in general, higher temperatures cause a drastic metabolic depression that is evident in tests performed between 15 and 20°C. This depression is highest in individuals from the lowest elevation site in the study (963 m), suggesting that this species complex is physiologically constrained to high elevations (Bernardo and Spotila 2006).

3.3. Temperature and chytridiomycosis as an example of synergistic environmental effects

Some authors defend the idea that global warming is a key factor promoting the action of B. dendrobatidis (Pounds et al. 2006a, Bosch et al. 2007), while admitting that this pathogen, even if present, may not always be a main causative agent of decline (Pounds et al. 2006b). Although the available studies show association more than causation, they point to a potential crucial interaction in anuran conservation. Comparative physiology is already helping to elucidate the relationship between chytridiomycosis and temperature. It is clear that the relationships between infectious agents and anuran hosts is influenced by thermoregulatory behaviors and by thermal ecology (Berger et al. 1998, Alford and Richards 1999). The varieties of the fungus B. dendrobatidis studied so far grow best at moderate temperatures, i.e., slightly above 20°C (Berger et al. 2004, Daszak et al. 2003, Burrowes et al. 2004, Pounds et al. 2006a), so that, global warming leading to increased daily body temperatures may increase the risk of infection of certain populations (Pounds et al. 2006a). However, temperatures well above the peak for fungal growth yet moderate for anurans (e.g., 28-30°C) inhibit or even kill the pathogen and help frogs to fight infection (Woodhams et al. 2003, Pounds et al. 2006a). Such temperatures can be attained by anurans through thermoregulatory behaviors leading to body temperatures higher than normal. Such thermoregulatory shifts were first reported for small thermophilic lizards (Kluger et al. 1975) but are currently known to occur in diverse vertebrate tetrapods including anurans, and to help animals to survive bacterial infection (Myhre et al. 1977, Sherman et al. 1991, Woodhams et al. 2003). Because opportunity for thermoregulation and typical body temperatures vary among anurans, even when comparing syntopic species (Navas 1996b), vulnerability to pathogens may vary as a consequence of differences in patterns of activity and preferred microhabitat. The relevance of thermal ecology to pathogen-host relationships in anurans is illustrated by studies on seasonality and environmental temperatures. During the winter months (Berger *et al.* 2004), or in environments likely to have summer temperatures lower than 30° C (Drew *et al.* 2006), the incidence of chytridiomycosis in Australian frogs increases. What we conclude is that a comprehensive analysis of microhabitat selection, patterns of activity, and field body temperatures is fundamental to fully understand anuran vulnerability to chytridiomycosis.

3.4. Pollutants

Because of its magnitude and ubiquity, a main factor threatening biodiversity in general, and amphibians in particular, is chemical pollution (Koeman 1991, Carey and Bryant 1995, Schiesari et al. 2007). The effects of pollutants on organisms in natural environments is the primary subject of ecotoxicology, whereas understanding the mechanisms by which organisms rid themselves of toxic substances is one topic of study in comparative physiology. These two fields meet in ecophysiological approaches focused on conservation biology, aiming to evaluate and predict the environmental risks of pollution contaminants to the environment (Koeman 1991, Stevenson et al. 2005). Because of a life-history usually involving aquatic larvae and terrestrial adults, and to a permeable and vascularized skin, amphibians seem particularly prone to uptake pollutants (Donnelly 1998, Maxell 2000, Schiesari et al. 2007). Although important caveats have been noted (Beebee and Griffiths 2005), amphibians have become widely recognized as indicators of environmental quality. They are lethally affected by petroleum products (Mahaney 1994), acidification (Rowe et al. 1992, Sadinski and Dunson 1992), insecticides (Berrill et al. 1993, 1994), and various metals (Freda 1991). Anuran species, however, differ in their ability to tolerate chemical contaminants (Hall and Henry 1992, Bridges and Semlitsch 2000, Rowe *et al.* 2001, Christin *et al.* 2004). These differences relate to the concept of allostatic capacity, and could be better understood using integrative approaches involving comparative physiology and ecotoxicology.

As evident from the discussion in Section 3.3, conservation physiology can contribute to understanding synergistic effects of environmental stressors. Pollutants may increase anuran vulnerability to pathogens, predators, ultraviolet radiation, or climate change (Little et al. 2000, Carey et al. 2001, Relyea and Millis 2001). One complication is that the effects of pollutants on organisms may be drastic only after chronic exposure. These types of effects, usually termed sublethal, include depressed disease resistance, inhibition of growth and development, decreased reproductive ability, inhibition of predator avoidance behaviors, and increased likelihood of developing morphological abnormalities (Lefcort et al. 1998, Maxell 2000, Christin et al. 2004, Gurushankara et al. 2007). Anuran populations are not exceptional, and may be affected sublethally by progressive accumulation of organic compounds or heavy metals in their tissues (Hall and Mulhern 1984). The accumulation of toxic substances sometimes results in morphological or physiological changes (Calow 1991, Alvarez et al. 1995, Rowe et al. 1996, 1998), and may affect energetics. In a number of animal species, the sublethal effects of toxic compounds include an increase in metabolic rate (Calow 1991, Hopkins et al. 1998, Beyers et al. 1999, Naab et al. 2001, Barbieri et al. 2002), but the effects of contaminants on anuran energetics are not well understood. The concentration of coal-ashderived pollutants is related to elevated metabolic rates in the bullfrog, Lithobates catesbeianus (Rowe et al. 1998), but not in Anaxyrus terrestris (Rowe et al. 2001). As in other examples cited before, the same kind and magnitude of environmental change, in this case pollutant type and concentration, may affect amphibian species in diverse, even contrasting ways (Calow and Sibly 1990, Koeman 1991). A full understanding of the relationship between anuran ecology, physiology, and ecotoxicology requires much additional work and information about the concentration, persistence, and spatiotemporal variation of pollutants in the environment (Davidson *et al.* 2002, Schiesari *et al.* 2007).

Although comparative physiology can help to clarify how anuran populations are affected by pollutants, the biogeographic zones with highest amphibian diversity have been neglected in these types of studies (Schiesari et al. 2007). Common contaminants such as organochlorine pesticides disrupt normal endocrine function and lead to reduced oogenesis and reproductive output in Xenopus laevis (Pickford and Morris 2003). Pesticides may also disrupt the normal course of steroid hormone production and result in demasculinization and hermaphroditism (Hayes et al. 2002, 2003). The threatened California red legged frog (Rana draytonii), at some point believed to be mainly affected by global warming, turned out to be particularly sensitive to agrochemicals (Davidson et al. 2001). Comparative immunological studies demonstrate that a realistic (i.e., likely to appear in the province of Quebec, Canada) mixture of agricultural pesticides reduce, although in different manners, the efficacy of the immune system of X. laevis and Lithobates pipiens (Christin et al. 2004). A mixture of pesticides used in US cornfields cause similar effects, and are associated with an increase in plasma levels of the stress hormone corticosterone (see note on stress hormones in Section 3.1) (Hayes et al. 2006). Comparative physiology has also helped to elucidate mechanisms behind specific pathological conditions of anurans exposed to pollution. For example, exposure to an acidic environment favors the opportunistic infection of the spleen by otherwise non-pathological bacteria, apparently by disrupting the integrity of epithelial barriers that normally prevent migration of intestine bacteria to the blood (Simon et al. 2002).

3.5. Habitat fragmentation

A consequence of urban and agricultural development is the transformation of formerly large extensions of continuous habitat into isolated patches of preserved areas (Miller and Cale 2000, Andersen et al. 2004, Pinto and Brito 2005). Habitat fragmentation may benefit a number of open-area anurans (Carnaval 2002), for example Chaunus marinus and Scinax ruber, two species frequently observed on the roads and farms built in the Amazonian forest (Duellman 1999). Local extinction and reduction of species richness, however, is a more common trend in fragmented habitats (Connor and McCoy 1979, Laan and Verboom 1990, Fahrig and Merriam 1994, Zuidema et al. 1996, Marsh and Pearman 1997, Vallan 2000, Pineda and Halffter 2004, Drinnan 2005, Funk et al. 2005, Bell and Donnelly 2006, Neckel-Oliveira and Gascon 2006). The anuran communities of fragmented habitats normally exhibit a subgroup of the presumed original communities (Ficetola and De Bernardi 2004, Bell and Donnelly 2006, Woinarski et al. 2006), and why this is so is a fundamental question for anuran conservation.

The effects of habitat fragmentation on anuran communities probably has three main groups of causes. The first one is reduced arthropod diversity (Maleque et al. 2006), and the possibility that an impoverished diet makes anuran more fragile or susceptible to infection (see Section 3.1). The second group of causes relates to reproduction, a topic mentioned already in the discussion of pollution and global climate change, and that we will treat in the next paragraph from the perspective of ability to reach breeding grounds. The third group of causes refers to the pool of changes in the magnitude and variance of physical variables that characterize habitat fragmentation. Habitat fragments are, from many points of view, poorer environments than the original habitats, and are also more prone to extremes in the sense that a number of relevant physical characteristics

become less predictable. Compared to the core of the forest, for example, the edges of forest patches are more exposed to wind, solar radiation, thermal variance and dehydration. Not only do these characteristics reduce the effective habitat area available for forest-adapted organisms (Saunders et al. 1991, Murcia 1995), but habitat remnants are more exposed to agrochemical inputs, influence from logging, livestock grazing, and induced fire (Turner 1996). The ability of anuran species to tolerate the novel physical regimes brought about by fragmentation relates to the concept of allostatic capacity cited in Section 2 and applies to all developmental phases, from eggs to adults. Although tadpoles of some species tolerate a wide range of water conditions in modified habitats (Loman and Lardner 2006), specific phases of development may be bottlenecks in modified habitats (see Section 3.4).

One correlate of forest fragmentation that may have profound effects on anuran communities is decreased opportunity for reproduction (Zimmerman and Bierregaard 1986). Forest anurans have complex and diverse modes of reproduction (Donnelly and Guyer 1994), but many species can be coarsely classified as pond-breeders and litter dwellers, the latter referring to those species with modes of reproduction that do not depend on sources of standing water (Donnelly 1998). Small forest fragments may not contain bodies of water proper for reproduction, and thus force individuals, perhaps even on a daily basis, to travel back and forth to breeding grounds. Doing so generates a physiological challenge whose magnitude relates to the conditions of the open areas surrounding fragments, particularly water availability and temperature, and to the distance between forest fragments and water bodies. How challenging a specific site is, then, depends on landscape ecology, mainly distance among patches (Brown and Kodric-Brown 1977), characteristics of the environment surrounding them (Fahrig and Merriam 1994, Gascon et al. 1999), and the number of corridors connecting patches (Lima and Gascon 1999).

It is generally believed that the threat of habitat fragmentation for anuran amphibians is worsened because of site fidelity and limited ability for dispersal (Blaustein et al. 1994, Marsh and Pearman 1997, Smith and Green 2005). The community of pond-breeding anurans in small fragments is likely to be composed mainly by the species with better ability to reach breeding grounds and circulate among neighboring fragments (Becker et al. 2007), which is a partial function of body size and metabolic physiology. The body of literature dedicated to the physiology of anuran locomotion shows dramatic differences among species in power and stamina that relate to the biochemical profile of leg muscle fibers (Taigen et al. 1982, Taigen and Pough 1985, Pough and Taigen 1990). Because these and other traits of exercise physiology determine the ability for locomotion of anurans, they should influence also the genetic flow among anuran populations isolated in forest fragments, independently of their mode of reproduction. Despite the importance of understanding whether exercise physiology influences amphibian vulnerability to habitat fragmentation, we were unable to detect studies carried with this purpose.

3.6. Exotic species

Human activity has been historically associated with the deliberate or accidental transport of animal and plant species outside their natural ranges, and to related changes in local patterns of biodiversity (Lonsdale 1999, Mack *et al.* 2000, McKinney 2002a, b). Exotic species may affect local diversity through their interactions with native species, particularly when their introduction occurs concomitantly with human-related ecological deterioration (Sax and Gaines 2003, Williamson 1996, Lonsdale 1999). Tolerance to anthropic environments is a characteristic of two anuran species that are currently considered feral pests in several parts of the world, the cane toad (Chaunus marinus) and the Bullfrog (Lithobates catesbeianus). The bullfrog is the principal invasive alien species in Brazilian amphibian communities (Guix 1999, Fortes et al. 2004, Rocha-Miranda et al. 2006), a process that has been facilitated by artificial dispersal related to teaching and aquiculture (Guiz 1999, Rocha-Miranda et al. 2006). The first Brazilian bullfrog cultures were established in 1935 (ISSG, 2005), and feral populations in State of São Paulo were detected in 1988 (Bruneau and Magnin 1980, Bury and Whelan 1984). Although froglets feed mainly on insects (Minton 1949 apud Guix, 1999), adults are voracious predators of crustaceans and other small or young vertebrates, including other anurans (Guix 1999). Bullfrog tadpoles may feed on eggs and larvae of other anuran species (Ehrlich 1979), but tend not to be palatable to many local predators (Rocha-Miranda et al. 2006). Bullfrogs may also offer indirect risk to local anurans, as the colonization of natural areas by this species may contribute to the dispersal of disease, including chytridiomycosis (Mazzoni et al. 2003, Hanselmann et al. 2004). All these traits enhance the possible role of this invasive species as a threat to the local anuran fauna (Jim 1997, Guix 1999, Rocha-Miranda et al. 2006).

The success of feral species to colonize given environments appears to follow some broad ecological rules, but at the same time is case-specific. On one hand, invasion biologists have proved that the species richness of a community relates to the likelihood of invasion by exotic species (Stohlgren et al. 1999, Meiners et al. 2004, Chown et al., 2005). Studies in the Southern Ocean Islands showed that energy flow into the islands relates positively to the diversity of both indigenous and exotic species. This appears to be so because favorable local energetic conditions, which are in turn associated with greater niche partitioning, favor diversity independently of whether the components of modified communities are native or exotic (Chown et al. 2005). On the other hand, some exotic species are far more likely than others to dominate new settings to the point of changing ecosystem structure and functioning (D'Antonio and Dudley 1995, Mack *et al.* 2000).

Why some exotic species have the physiological and behavioral ability to outcompete local species is a question that lies in the interface between physiology and invasion biology. From one point of view, some successful invasive species may have traits of physiology that make them particularly competitive in new environments. These may relate to reproductive output and growth rates, but also to subtle aspects of physiology. For example, tadpoles of L. catesbeianus can recognize and react to cues of novel predators, and this neural ability may favor its capacity to colonize novel settings (Pearl et al. 2003). It is also possible that the ability of species to gather and process energy affects the results of ecological interactions. One case study that has received particular attention is the interaction between introduced bullfrogs on populations of the native California red-legged frog Rana aurora. Although bullfrogs usually cause declines of Rana aurora, the negative effects of this introduced species are more dramatic when food resources are clumped and almost negligible when resources are scattered through the ponds. One conclusion is that the mechanisms by which exotic anuran species affect native frogs need not to be direct or obvious (Adams 2000); furthermore, such effects may include reaction norms of physiological processes. Tadpoles of native species may respond with different shifts in level of activity and growth rates to the presence of L. catesbeianus tadpoles and these differences may affect the competitive interactions with native and introduced species (Monello et al. 2006).

4. The Brazilian Scenario

The global phenomenon of decline and extinction of amphibian populations has not

spared Brazil (Heyer et al. 1988, 1990, Weygoldt 1989, Haddad and Sazima 1992, Bertoluci and Heyer 1995, Young et al. 2001, Eterovick et al. 2005), and is evident even in protected areas regularly visited since the 1970s (Heyer et al. 1988, Weygoldt 1989, Bertoluci and Heyer 1995, Pombal and Haddad 1999, Eterovick et al. 2005). The reasons for the decline of Brazilian anurans are largely unknown, but possible causative agents include habitat fragmentation, pollutants, infections, climatic changes, invasion by feral species, wildlife trade (Silvano and Segalla 2005), and possible synergistic effects among some or all these factors (Young et al. 2001). The climatic conditions of Brazil offer large extensions suitable for the action of the pathogen fungus B. dendrobatidis (Carnaval et al. 2006, Toledo et al. 2006a, b) and 23 species have been reported as infected by this fungus in the Brazilian Atlantic forest (Carnaval et al. 2006, Toledo et al. 2006a, b). Scientists speculate about a possible expansion to the Cerrado and the Pantanal (Ron 2005, Toledo et al. 2006a, b), but little is known about the current distribution of B. dendrobatidis in Brazil and its possible consequences to local species (Toledo et al. 2006a, b). Discussing cause-effect relationships associated with declines of Brazilian anurans is difficult because of inadequate information about natural history and ecology, particularly in terms of long-term monitoring (Silvano and Segalla 2005). This situation is further complicated by the size of the country, the diversity of its anuran fauna (Silvano and Segalla 2005), and the meager information available on the physiological ecology of Brazilian anurans. It is symptomatic of these problems that we were unable to find records about effects of common toxic compounds, such as pesticides, on Brazilian amphibian species. The absence of data, in this case, supports the claim by Schiesari et al. (2007), that areas with high amphibian diversity have received very little attention regarding the consequences of pollutant spill (see Section 3.5).

The problem of habitat fragmentation is well illustrated in Brazil by the natural history of Atlantic forest, although other biomes such as the Cerrado also deserve attention (Brasileiro et al. 2005). The Atlantic forest is the richest Brazilian biome in terms of anuran fauna, and shelters more than 300 species, many of which are endemic (Duellman 1999). During the past three decades, this biome has been fragmented and altered (Pinto and Brito 2005) to a point where only about 5% of the original area remains as fragments of diverse sizes (SOS Mata Atlântica, 1998). The fragmentation of the Atlantic forest may be less stressing for species that do not require ponds for reproduction, such as members of the genus Eleutherodactylus (Brachycephalidae), but is likely to affect species requiring ponds or streams (Heyer et al. 1990, Becker et al. 2007). When bodies of water are excluded from Atlantic forest fragments, anurans that persist because of either their ability to tolerate the ecological conditions of gaps, or their capacity for locomotion, may have an advantage (see Section 3.5). Although we defend emphatically the importance of studies aiming to identify the ecological outcome of fragmentation for anurans of the Atlantic forest, a necessary extension in scope requires understanding why some forest species tolerate the physical changes associated with fragmentation better than others. For example, comparative studies of the physiology of species adapted to natural forest gaps and inside forest may help in understanding which species, and why species are affected by deforestation (Haddad et al. 2007). Overall, animal physiological adaptation to the Atlantic forest has been largely overlooked despite its importance for conservation (Navas et al. 2007b).

The effects of climate change in Brazil have been discussed mainly in terms of transformation of lands for agriculture (Cerri *et al.* 2007), but recent efforts have been devoted to understanding comprehensive effects on biological diversity (Marengo 2006). As

discussed in Section 3.2, global warming and extreme climatic events run in parallel, so it is worth asking to which extent atypical climatic events may have already influenced the anuran fauna of Brazil. According to Heyer et al. (1988), a severe frost may have had a dramatic effect on the anuran communities of Southeastern Brazilian Atlantic forest. From data spanning 35 years, Heyer and collaborators observed drastic declines in the populations of Hyalinobatrachium eurygnathum, Scinax perpusillus, Leptodactylus marmoratus, Eleutherodactylus guentheri, E. parvus, and Hylodes phyllodes populations, and also registered the extinction of Crossodactylus dispar, Cycloramphus boraceiensis, C. semipalmatus, Hylodes asperus and Thoropa taophora. The reported population crashes coincided with a four-day long frost that occurred in 1979 during which temperatures dropped to -2.6°C. Whereas some species that were decimated in 1979 (Heyer et al. 1988) recovered as to be considered abundant again (S. perpusilla and Hylodes phyllodes; Bertoluci and Heyer 1995), others disappeared (C. dispar, C. gaudichaudii, C. boraciencis, C. semipalmatus, H. asperus and T. taophora; Bertoluci and Heyer 1995). Other cases of population declines in Brazilian frogs have been attributed to unusually dry winters (Weygoldt 1989); it seems clear that short-term but extreme climatic events may become part of the evolutionary history of anurans and affect their distribution (Vanzolini and Williams 1970, Heyer et al. 1988). Still, we must ask why some species, among the many equally exposed to such extremes through time, are particularly sensitive to cold. Hot temperatures may also affect Brazilian anurans, particularly those in semi-arid habitats (see Section 3.2). Postmetamorphic toads from the Brazilian Caatinga, a hot semiarid biome in Northeastern Brazil, are diurnal and disperse by hopping over long distances on hot substrates. These toads exhibit the highest thermal tolerance reported for anurans, yet are active at conditions close to their thermal tolerances (Navas *et al.* 2007a). They are unlikely to tolerate the 2-5°C increase in mean temperature, and the concomitant decrease in humidity, predicted for the Caatinga at the end of the 21st century (Marengo 2006).

It is worth noting that some anuran species, native or invasive, benefit from environmental changes resulting from human activities. For example, anurans that lay eggs in still water, or that lay eggs protected by foam nests, may be more tolerant to regional temperature shifts (Haddad and Prado 2005, Haddad et al. 2007). As far as such bodies of water are seasonally available, these species may occupy areas otherwise becoming more arid (see Haddad et al., 2007). Bullfrogs are very competitive in warm and open ponds and by the side of rivers in modified environments, which are common in Brazil. Asking why bullfrogs are favored in areas exposed to human impact; whether they are, as it seems, less susceptible to pollutants than most native species, or whether their tadpoles are physiologically more capable than those of native species, may bring new insights to explain the success of this exotic species. Reproductive output is an issue too because in comparison with individuals within the natural range, Brazilian bullfrogs exhibit longer reproductive periods and higher growth rates (Fortes et al. 2004). These shifts in natural history may be just the passive consequence of exposure to warmer conditions and the absence of a winter, but they may reflect as well adjustments in the thermal biology of the species to tropical conditions. Brazilian bullfrogs, and alien species in general, constitute a valuable resource to understanding why some species are more successful at invading new areas than others, and how the patterns of relative success might change as both local and regional environments change (Frenot et al. 2005).

The conservation of Brazilian anurans would benefit from studies using a number of techniques that, although quite restricted in the country, have proven useful to monitor or predict physiological shifts along ecological gradients. These include 1) the combination of physiological research with biophysical modeling to identify how large-scale climatic variables may affect individual organisms (Spotila et al. 1992, Bartelt and Peterson 2005, Helmuth et al. 2005), 2) data logging to monitor physiological responses to environmental variables (Block 2005), and 3) integration of studies in physiological ecology with data coming from Geographic Information Systems (GIS). Spatial data from GIS allow graphical display and analysis at a large scale, and have proven valuable to understand how human activity affects animal energetics and ecological success in disturbed environments (Porter et al. 2000). As examples, combined data from GIS and experimental physiology have been used to predict the areas in which the conditions of humidity and temperature are appropriate for successful hatching in a lizard species (Porter et al. 2000), and the potential distribution of introduced species such as Chaunus marinus in Australia (Sutherst et al. 1996).

5. Conclusions

As previous authors have pointed out (Blaustein et al. 2002), it is often difficult to assign a single cause to the decline of an anuran population because some noxious factors act in the long term, and because synergistic interaction among environmental variables obscure individual effects. Independent of these problems, the examples of environmental change highlighted in this review would benefit by further studies in thermal biology, water balance, exercise physiology, thermal physiology, endocrinology, immunology, and metabolic physiology. Studies in physiological ecology are of remarkable practical value to understand the susceptibility of anurans to environmental extremes, including retrospective studies where preserved specimens and climate records are available. Comparative physiology would help to explain the underlying mechanisms supporting these differences among species, and evolutionary physiology could elucidate the scope and constraints of possible adaptive change. All these topics and approaches converge at conservation physiology, and may generate important applicable information to understand anuran declines and improve management decisions.

One conclusion emerging from our analysis is the need for integration, and the potential value of studies involving cooperation between ecologists and physiologists. Despite the huge body of literature on anuran thermal biology, for example, it is still difficult to assess how changes in climate may affect the overall reproductive output, particularly in species that hibernate or aestivate. Physiological ecology associated with habitat fragmentation is also a very constructive field of research. By investigating the environmental conditions of the gaps and the physiological tolerance of anuran species to these conditions we could answer questions such as how far or for how long can individual anurans successfully progress into the gaps between habitat fragments? To what extent do basic corridors that improve shade and humidity help to connect isolated forest populations? Although it is clear that habitat loss and fragmentation affect amphibian performance, survival, and reproductive success, we need to understand why in order to improve management. Interdisciplinary studies may also help to elucidate complex patterns, for example possible interaction between fragment area, arthropod diversity, diet, skin peptides, and microbial defense in anurans.

Brazil is already well-prepared with infrastructure, databases, and ecological background to make anuran conservation physiology a particularly successful enterprise. An additional advantage is the diverse geography and large extension of the country, given that species distributed along significant ecological gradients are of special value to predict the types of changes that would occur because of environmental change such as global warming (Miles 1994, Donnelly 1998). Brazil also has the potential to support studies aiming to elucidate which combinations of ecophysiological factors (e.g., activity temperature, dependence on soil moisture, and skin permeability) affect the likelihood of a chytridiomycotic infection, what are the consequences of global climate change to anurans, and why various specific fractions of the original community survive in forest fragments of different sizes. These and many other questions in conservation physiology can readily be converted into testable hypotheses that would greatly contribute to conservation and management of Brazilian anurans.

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References

- Adams, M. J. 2000. Pond permanence and the effects of exotic vertebrates on anurans. *Ecological Applications* 10: 559–568.
- Alford, R. A. and S. J. Richards. 1999. Global amphibian declines: a problem in applied ecology. *Annual Review of Ecology and Systematics* 30: 133–165.
- Alvarez, R., M. P. Honrubia and M. P. Herraez. 1995. Skeletal malformations induced by the insecticides ZZ-Aphox and folidol during larval development of *Rana* perezi. Archives of Environmental Contamination and Toxicology 28: 349–356.
- Andersen, L. W., K. Fog and C. Damgaard. 2004. Habitat fragmentation causes bottlenecks and inbreeding in the European tree frog (*Hyla arborea*). Proceedings of Royal Society B 271: 1293–1302.
- Angilletta, M. J., P. E. Niewiarowski and C. A. Navas. 2002. The evolution of thermal physiology in ectotherms. *Journal of Thermal Biology* 27: 249–268.
- Angilletta, M. J., R. S.Wilson, C. A. Navas, and R. S. James. 2003. Tradeoffs and the evolution of thermal reaction norms. *Trends in Ecology and Evolution 18*: 234–240.

- Ashcroft, J. W., Z. B. Zalinger, C. R, Bevier, and F. A. Fekete. 2007. Antimicrobial properties of two purified skin peptides from the mink frog (*Rana* septentrionalis) against bacteria isolated from the natural habitat. Comparative Biochemistry and Physiology (in press).
- Barbieri, E., I. R. Oliveira and P. C. S. Serralheiro. 2002. The use of metabolism to evaluate the toxicity of dodecil benzene sodium sulfonate (LAS-C12) on the *Mugil platanus* (mullet) according to the temperature and salinity. *Journal of Experimental Marine Biology* and Ecology 277: 109–127.
- Bartelt, P. E. and C. R. Peterson. 2005. Physically modeling operative temperatures and evaporation rates in amphibians. *Journal of Thermal Biology* 30: 93– 102.
- Beardall, J., S. Beer and J. A. Raven. 1998. Biodiversity of marine plants in an era of climate change: Some predictions based on physiological performance. *Botanica Marina 41*: 113–123.
- Becker, C. G., C. R. Fonseca, C. F. B. Haddad, R. F. Batista, and P. I. Prado. 2007. Habitat split and the global decline of amphibians. *Science 318*: 1775– 1776.
- Beebee, T. J. C. 1995. Amphibian breeding and climate. Nature 374: 219–220.
- Beebee, T. J. C. 2002. Amphibian phenology and climate change. *Conservation Biology* 16: 1454–1454.
- Beebee, T. J. C. and R. A. Griffiths. 2005. The amphibian decline crisis: A watershed for conservation biology? *Biological Conservation 125*: 271–285.
- Beecher, N. A. and G. E. Demas. 2004. Frog declines: Exploring connections between climate change, immunity and disease. *Integrative and Comparative Biology* 44: 522–522.
- Bell, K. E. and M. A. Donnelly. 2006. Influence of forest fragmentation on community structure of frogs and lizards in Northeastern Costa Rica. *Conservation Biology* 20: 1750–1760.
- Berger, L., R. Speare, P. Daszak, D. E. Green, A. A. Cunningham, C. L. Goggin, R. Slocombe, M. A. Ragan, A. D. Hyatt, K. R. McDonald, H. B. Hines, K. R. Lips, G. Marantelli, and H. Parkes. 1998. Chytridiomycosis causes amphibian mortality associated with population declines in the rain forests of Australia and Central America. Proceedings of the National Academy of Sciences of the United States of America 95: 9031–9036.
- Berger, L., R. Speare, H. B. Hines, G. Marantelli, A. D. Hyatt, K. R. McDonald, L. F. Skerratt, V. Olsen, J. M. Clarke, G. Gillespie, M. Mahony, N. Sheppard, C. Williams, and M. J. Tyler. 2004. Effect of season and temperature on mortality in amphibians due to chytridiomycosis. Australian Veterinary Journal 82: 434–439.

- Bernardo, J. and J. R. Spotila. 2006. Physiological constraints on organismal response to global warming: mechanistic insights from clinally varying populations and implications for assessing endangerment. *Biology Letters* 2: 135–139.
- Berrill, M., S. Bertram, L. McGillivray, M. Kolohon, and B. Pauli. 1994. Effects of low concentrations of forestuse pesticides on frog embryos and tadpoles. *Environmental Toxicology and Chemistry* 13: 657– 664.
- Berrill, M., S. Bertram, A. Wilson, S. Louis, D. Brigham, and C. Stromberg. 1993. Lethal and sublethal impacts of pyrethroid insecticides on amphibian embryos and tadpoles. *Environmental Toxicology and Chemistry 12*: 525–539.
- Bertoluci, J. and W. R. Heyer. 1995. Boracéia update. Froglog 14: 2–3.
- Berven, K. A. 1982. The genetic basis of altitudinal variation in the woodfrog *Rana sylvatica*: 1 An experimental analysis of life history traits. *Evolution* 36: 962–983.
- Beyers, D. W., J. A. Rice, W. H. Clements, and C. J. Henry. 1999. Estimating physiological cost of chemical exposure: integrating energetics and stress to quantify toxic effects in fish. *Canadian Journal of Fisheries* and Aquatic Sciences 56: 814–822.
- Blaustein, A. R. and J. M. Kiesecker. 2002. Complexity in conservation: lessons from the global decline of amphibian populations. *Ecology Letters* 5: 597–608.
- Blaustein, A. R., D. B. Wake and W. P. Sousa. 1994. Amphibian declines: Judging stability, persistence, and susceptibility of populations to local and global extinctions. *Conservation Biology* 8: 60–71.
- Blaustein, A. R., L. K. Belden, D. H. Olson, D. M. Green, T. L. Root, and J. M. Kiesecker. 2001. Amphibian breeding and climate change. *Conservation Biology* 15: 1804–1809.
- Blaustein, A. R., T. L. Root, J. M. Kiesecker, L. K. Belden, D. H. Olson, and D. M. Green. 2003. Amphibian breeding and climate change: Reply to Corn. *Conservation Biology* 17: 626–627.
- Block, B. A. 2005. Physiological ecology in the 21st century: Advancements in biologging science. *Integrative and Comparative Biology* 45: 305–320.
- Bosch, J., L. M. Carrascal, L. Duran, S. Walker, and M. C. Fisher. 2007. Climate change and outbreaks of amphibian chytridiomycosis in a montane area of Central Spain; is there a link? *Proceedings of the Royal Society B-Biological Sciences* 274: 253–260.
- Brasileiro, C. A., R. J. Sawaya, M. C. Kiefer, and M. Martins. 2005. Amphibians of an open cerrado fragment in southeastern Brazil. *Biota Neotropica* 5: 93–109.

- Brauner, C. J., D. J. Randall, J. F. Neuman, and R. V. Thurston. 1994. The effect of exposure to 1,2,4,5-tetrachlorobenzene and the relationship between toxicant and oxygen uptake in rainbow trout. *Environmental Toxicology and Chemistry* 13: 1813–1820.
- Bridges, C. M. and R. D. Semlitsch. 2000. Variation in pesticide tolerance of tadpoles among and within species of ranidae and patterns of amphibian decline. *Conservation Biology* 14: 1490–1499.
- Brown, J. H. and A. Kodric-Brown. 1977. Turnover rates in insular biogeography: effect of immigration on extinction. *Ecology* 58: 445–449.
- Bruneau, M. and E. Magnin. 1980. Croissance, nutrition et reproduction des ouaouarons *Rana catesbeiana* Shawn (Amphibia: Anura) des Laurentides au nord de Montréal. *Canadian Journal of Zoology* 58: 175–183.
- Burrowes, P. A., R. L. Joglar and D. E. Green. 2004. Potential causes for amphibian declines in Puerto Rico. *Herpetologica* 60: 141–154.
- Bury, R. B. and J. A. Whelan. 1984. Ecology and management of the bullfrog. U.S. Department of the Interior, Fish and Wildlife Service, Resource Publication 155, Washington, D.C.
- Calow, P. 1991. Physiological costs of combating chemical toxicants: ecological implications. *Comparative Biochemistry and Physiology C 100*: 3–6.
- Calow, P. and R. M. Sibly. 1990. A physiological basis of population processes: ecotoxicological implementations. *Functional Ecology* 4: 283–288.
- Carey, C. 2005. How physiological methods and concepts can be useful in conservation biology. *Integrative and Comparative Biology* 45: 4–11.
- Carey, C. and C. J. Bryant. 1995. Possible interrelations among environmental toxicants, amphibian development, and decline of amphibian populations. *Environmental Health Perspectives 103*: 13–17.
- Carey, C., W. R. Heyer, J. Wilkinson, R. A. Alford, J. W. Arntzen, T. Halliday, L. Hungerford, K. R. Lips, E. M. Middleton, S. T. Orchard, and A. S. Rand. 2001. Amphibian declines and environmental change: Use of remote-sensing data to identify environmental correlates. *Conservation Biology* 15: 903-915.
- Carnaval, A. C. O. Q. 2002. Phylogeography of four frog species in forest fragments of northeastern Brazil – a preliminary study. *Integrative and Comparative Biology* 42: 913–921.
- Carnaval, A. C. O. Q., R. Puschendorf, O. L. Peixoto, V. K. Verdade, and M. T. Rodrigues. 2006. Amphibian chytrid fungus broadly distributed in the Brazilian Atlantic Rain Forest. *Ecohealth* 3: 41–48.
- Cerri, C. E. P., G. Sparovek, M. Bernoux, W. E. Easterling, J. M. Melillo, and C. C. Cerri. 2007. Tropical agricul-

ture and global warming: impacts and mitigation options. *Scientia Agricola* 64: 83–99.

- Christin, M. S., L. Menard, A. D. Gendron, S. Ruby, D. Cyr, D. J. Marcogliese, L. Rollins-Smith, and M. Fournier. 2004. Effects of agricultural pesticides on the immune system of *Xenopus laevis* and *Rana pipiens*. *Aquatic Toxicology* 67: 33–43.
- Chown, S. L., B. Hull and K. J. Gaston. 2005. Human impacts, energy availability and invasion across Southern Ocean Islands. *Global Ecology and Biogeography* 14: 521–528.
- Collins, J. P. 2003. Global amphibian declines: sorting the hypotheses. *Diversity and Distributions* 9: 89–98.
- Connor, E. F. and E. D. McCoy. 1979. The statistics and biology of the species-area relationship. *The American Naturalist* 113: 833.
- Corn, P. S. 2003. Amphibian breeding and climate change: Importance of snow in the mountains. *Conservation Biology* 17: 622–625.
- D'Antonio, C. M. and T. L. Dudley. 1995. Biological invasions as agents of change on islands versus mainlands. Pp. 103–121 in P. M. Vitousek, L. L. Loope and H. Andersen (eds.), *Islands: biological diversity* and ecosystem function. Berlin, Springer.
- D'Cruz, L. M., J. J. Dockray, I. J. Morgan, and C. M. Wood. 1998. Physiological effects of sublethal acid exposure in juvenile rainbow trout on a limited or unlimited ration during a simulated global warming scenario. *Physiological Zoology* 71: 359–376.
- Dale, V. H., S. M. Pearson, H. L. Offerman, and R. V. Oneill. 1994. Relating patterns of land-use change to faunal biodiversity in the Central Amazon. *Conservation Biology* 8: 1027–1036.
- Daly, J. W. 1995. The chemistry of poisons in amphibian skin. Proceedings of the National Academy of Sciences of the United States of America 92: 9–13.
- Daszak, P., A. A. Cunningham and A. Hyatt. 2001. Anthropogenic environmental change and the emergence of infectious diseases in wildlife. Acta Tropica 78: 103–116.
- Daszak, P., A. A. Cunningham and A. Hyatt. 2003. Infectious disease and amphibian population declines. *Diversity and Distributions 9*: 141–150.
- Daszak, P., D. E. Scott, A. M. Kilpatrick, C. Faggioni, J. W. Gibbons, and D. Porter. 2005. Amphibian population declines at savannah river site are linked to climate, not chytridiomycosis. *Ecology* 86: 3232–3237.
- Davidson, C., H. B. Shaffer and M. Jennings. 2001. Declines of the California red-legged frog: Climate, UV-B, habitat, and pesticides hypotheses. *Ecological Applications 11*: 464–479.
- Davidson, C., H. B. Shaffer and M. Jennings. 2002. Spatial

tests of the pesticide drift, habitat destruction, UV-B and climate change hypotheses for California amphibian declines. *Conservation Biology* 16: 1588–1601.

- Donnelly, M. A. 1998. Potential effects of climate change on two neotropical amphibian assemblages. *Climatic Change* 39: 541–561.
- Donnelly, M. A. and C. Guyer. 1994. Patterns of reproduction and habitat use in an assemblage of neotropical hylid frogs. *Oecologia* 98: 291–302.
- Drew, A., E. J. Allen and L. J. S. Allen. 2006. Analysis of climatic and geographic factors affecting the presence of chytridiomycosis in Australia. *Diseases of Aquatic Organisms* 68: 245–250.
- Drinnan, I. N. 2005. The search for fragmentation thresholds in a Southern Sydney Suburb. *Biological Conservation* 124: 339–349.
- Duellman, W. E. 1999. Distribution patterns of amphibians in South America. Pp. 255–328 in W.E. Duellman (ed.), Patterns of Distribution of Amphibians: a global perspective. Baltimore, The John Hopkins University Press.
- Ehrlich, D. 1979. Predation by bullfrogs tadpoles (*Rana catesbeiana*) on eggs and newly hatched larvae of the plains leopard frog (*Rana blairi*). Bulletin of the Maryland Herpetology Society 15: 25–26.
- Eterovick, P. C., A. C. O. Q. Carnaval, D. M. Borges-Nojosa, D. L. Silvano, M. V. Segalla, and I. Sazima. 2005. Amphibian declines in Brazil: an overview. *Biotropica* 37: 166–179.
- Fahrig, L. and G. Merriam. 1994. Conservation of fragmented populations. *Conservation Biology* 8: 50–59.
- Ficetola, G. F. and F. De Bernardi. 2004. Amphibians in a human-dominated landscape: the community structure is related to habitat features and isolation. *Biological Conservation 119*: 219–230.
- Freda, J. 1991. The effects of aluminum and other metals on amphibians. *Environmental Pollution* 71: 305–328.
- Frenot, Y., S. L. Chown, J. Whinam, P. M. Selkirk, P. Convey, M. Skotnicki, and D. M. Bergstrom. 2005. Bipological invasions in the Antarctic: extent impacts and implications. *Biological Reviews* 80: 45–72.
- Fortes, V. B, E. M. L. Gonsales, A. Branco, F. C. Alves, and A.K. Martins. 2004. Perigo: antíbio exótico ameaça biodiversidade brasileira. *Revista Sul Ambiental 4*: 10.
- Funk, W. C., A. E. Greene, P. S. Corn, and F. W. Allendorf. 2005. High dispersal in a frog species suggests that it is vulnerable to habitat fragmentation. *Biology Letters* 1: 13–16.
- Gascon, C., T. E. Lovejoy, R. O. Bierregaard, J. R. Malcolm, P. C. Stouffer, H. L. Vasconcelos, W. F. Laurance, B. Zimmerman, M. Tocher, and S. Borges. 1999. Matrix habitat and species richness in tropical

forest remnants. *Biological Conservation* 91: 223–229.

- Guix, J. C. 1999. Introdução e colonização de Rana cateseiana Shaw, 1802 em um pequeno vale no município de Suzano (SP), sudeste do Brasil. Grupo de Estudos Ecológicos Série Documentos 2: 32–34.
- Gurushankara, H. P., S. V. Krishnamurthy and V. Vasudev. 2007. Effect of malathion on survival, growth, and food consumption of indian cricket frog (*Limnonectus limnocharis*) tadpoles. Archives of Environmental Contamination and Toxicology 52: 251–256.
- Haddad, C. F. B. and C. P. A. Prado. 2005. Reproductive modes in frogs and their unexpected diversity in the Atlantic forest of Brazil. *BioScience* 55: 207–217.
- Haddad, C. F. B. and I. Sazima. 1992. Anfíbios anuros da Serra do Japi. Pp. 188–211 in P. Morellato (ed.), História Natural da Serra do Japi: ecologia e preservação de uma área florestal no sudeste do Brasil. Campinas, Editora da UNICAMP.
- Haddad, C. B. F., J. G. Giovanelli and J. M. B. Alexandrino. 2007. O aquecimento global e seus efeitos na distrbuição e declínio dos anfíbios. In press in M. S. Buckeridge (ed.), A Biologia e as Mudanças Climáticas no Brasil.
- Hall, R. J. and P. F. P. Henry. 1992. Assessing effects of pesticides on amphibians and reptiles: status and needs. *Journal of Herpetology* 2: 65–71.
- Hall, R. J. and B. M. Mulhern. 1984. Are anurans heavy metal accumulators? Pp. 123–133 in R. A. Seigel, L. E. Hunt, L. M. Knight, N. L. Zuschlag (eds.), Vertebrate Ecology and Systematics - a Tribute to Henry S. Fitch. Lawrence, Museum of Natural History, University of Kansas.
- Hanselmann, R., A. Rodríguez, M. Lampo, L. Fajardo-Ramos, A. Aguirre, A. M. Kilpatrick, J. P. Rodríguez, and P. Daszak. 2004. Presence of an emerging pathogen of amphibians in introduced bullfrogs *Rana catesbeiana* in Venezuela. *Biological Conservation* 120: 115–119.
- Hayes, T. B., K. Haston, M. Tsui, A. Hoang, C. Haeffele, and A. Vonk. 2003. Atrazine-induced hermaphroditism at 0.1 ppb in American leopard frogs (*Rana pipiens*): laboratory and field evidence. *Environmental Health Perspectives 111*: 568–575.
- Hayes, T. B., A. Collins, M. Lee, M. Mendoza, N. Noriega, A. A. Stuart, and A. Vonk. 2002. Hermaphroditic, demasculanized frogs after exposure to the herbicide atrazine at low ecologically relevant doses. Proceedings of the National Academy of Sciences of the United States of America 99: 5476-5480.
- Hayes, T. B., P. Case, S. Chui, D. Chung, C. Haeffele, K. Haston, M. Lee, V. P. Mai, Y. Marjuoa, J. Parker, and M. Tsui. 2006. Pesticide mixtures, endocrine

disruption, and amphibian declines: are we underestimating the impact? *Environmental Health Perspectives 114*: 40–50.

- Helmuth, B., J. G. Kingsolver and E. Carrington. 2005. Biophysics, physiological ecology, and climate change: Does mechanism matter? *Annual Review of Physiology* 67: 177–201.
- Heyer, W. R., A. S. Rand, C. A. G. Cruz, and O. L. Peixoto. 1988. Decimations, extinctions, and colonizations of frog populations in southeast Brazil and their evolutionary implications. *Biotropica* 20: 230–235.
- Heyer, W. R., A. S. Rand, C. A. G. Cruz, O. L. Peixoto, and C. E. Nelson. 1990. Frogs of Boracéia. Arquivos de Zoologia,. São Paulo 31: 231–410.
- Hopkins, W. A., M. T. Mendonça and J. D. Congdon. 1997. Increased circulating levels of testosterone and corticosterone in Southern toads, *Bufo terrestris*, exposed to coal combustion waste. *General and Comparative Endocrinology 108*: 237–246.
- Hopkins, W. A., M. T. Mendonça, C. L. Rowe, and J. D. Congdon. 1998. Elevated trace element concentrations in southern toads, *Bufo terrestris*, exposed to coal combustion wastes. *Archives of Environmental Contamination and Toxicology* 35: 325–329.
- ISSG. 2005. Invasive Species Specialist Group of The World Conservation Union. http://www.issg.org.
- Jim, J. 1997 O impacto da criação de rãs sobre o meio ambiente. Pp. 163–164 in IX Encontro Nacional de Ranicultura, II International Meeting on Frog Research and Technology, Santos. Anais Santos: ABETRA/ABC. 236.
- Johnson, M. L., L. Berger, L. Philips, and R. Speare. 2003. Fungicidal effects of chemical disinfectants, UV light, desiccation and heat on the amphibian chytrid Batrachochytrium dendrobatidis. Diseases of Aquatic Organisms 57: 255–260.
- Karasov, W. H., R. E. Jung, S. Vanden Langenberg, and T. L. E. Bergeson. 2005. Field exposure of frog embryos and tadpoles along a pollution gradient in the Fox River and Green Bay ecosystem in Wisconsin, USA. Environmental Toxicology and Chemistry 24: 942–953.
- Karban, R. and S. Y. Strauss. 2004. Physiological tolerance, climate change, and a northward range shift in the spittlebug, *Philaenus spumarius*. Ecological Entomology 29: 251–254.
- Kluger, M. J., D. H. Ringler and M. R. Anver. 1975. Fever and survival. *Science* 188: 166–168.
- Koeman, J. H. 1991. From comparative physiology to toxicological risk assessment. *Comparative Biochemistry and Physiology C 100*: 7–10.
- Laan, R. and B. Verboom. 1990. Effects of pool size and isolation on amphibian communities. *Biological Conservation* 54: 251–262.

- Lefcort, H., R. A. Meguire, L. H. Wilson, and W. F. Ettinger. 1998. Heavy metals alter the survival, growth, metamorphosis, and antipredatory behavior of columbia spotted frog (*Rana luteiventris*) tadpoles. *Archives of Environmental Contamination and Toxicology 35*: 447–456.
- Lima, M. and C. Gascon. 1999. The conservation value of linear forest remnants in central Amazonia. *Biological Conservation* 91: 241–247.
- Lips, K. R., P. A. Burrowes and J. R. Mendelson J.R. III. 2005. Amphibian population declines in Latin America: a synthesis. *Biotropica* 37, 222–226.
- Lips, K. R., D. E. Green and R. Papendick. 2003a. Chytridiomycosis in wild frogs from southern Costa Rica. Journal of Herpetology 37: 215–218.
- Lips, K. R., J. D. Reeve and L. Witters. 2003b. Ecological traits predicting amphibian population declines in Central America. *Conservation Biology* 17: 1078–1088.
- Little, E. E., R. Calfee, L. Cleveland, R. Skinker, P. A. Zaga, and M. G. Barron. 2000. Photo-enhanced toxicity in amphibians: synergistic interactions of solar ultraviolet radiation and aquatic contaminants. *Journal* of the Iowa Academy of Science 107: 67–71.
- Loman, J. and B. Lardner. 2006. Does pond quality limit frogs *Rana arvalis* and *Rana temporaria* in agricultural landscapes? A field experiment. *Journal* of Applied Ecology 43: 690–700.
- Lonsdale, W. M. 1999. Global patterns of plant invasions and the concept of invasibility. *Ecology* 80: 1522– 1536.
- Mack, R. N., D. Simberloff, W. M. Lonsdale, H. Evans, M. Clout, and F. A. Bazzaz. 2000. Biotic invasions: causes, epidemiology, global consequences, and control. *Ecological Applications* 10: 689–710.
- Mahaney, P. A. 1994. Effects of freshwater petroleum contamination on amphibian hatching and metamorphosis. *Environmental Toxicology* 13: 259–265.
- Maleque, M. A., H. T. Ishii and K. Maeto. 2006. The use of arthropods as indicators of ecosystem integrity in forest management. *Journal of Forestry 104*: 113–117.
- Marengo, J. A. 2006. Mudanças climáticas globais e seus efeitos sobre a biodiversidade. Caracterização do clima atual e definição das alterações climáticas para o território brasileiro ao longo do século XXI. Ministério do Meio Ambiente.
- Marsh, D. M. and P. B. Pearman. 1997. Effects of habitat fragmentation on the abundance of two species of leptodactylid fogs in an Andean montane forest. *Conservation Biology* 11: 1323–1328.
- Matutte, B., K. B. Storey, F. C. Knoop, and J. M. Conlon. 2000. Introduction of synthesis of an antimicrobial peptide in the skin of the freeze-tolerant frog, *Rana* sylvatica, in response to environmental stimuli.

Federation of European Biochemical Societies Letters 483: 135–138.

- Maxell, B. A. 2000. Management of Montana's amphibians: a review of factors that may present a risk to population viability and accounts on the identification, distribution, taxonomy, habitat use, natural history, and the status and conservation of individual species. University of Montana, Wildlife Biology Program, Missoula.
- Mazzoni, R., A. C. Cunningham, P. Daszak, A. Apolo, E. Perdomo, and G. Speranza. 2003. Emerging pathogen of wild amphibians in frogs (*Rana catesbiana*) farmed for international trade. *Emerging Infectious Diseases* 9: 995–998.
- McEwen, B. S. and J. C. Wingfield. 2003. The concept of allostasis in biology and biomedicine. *Hormones and Behavior* 43: 2–15.
- McKinney, M. L. 2002a Do human activities raise species richness? Contrasting patterns in United States plants and fishes. *Global Ecology and Biogeography 11*: 343–348.
- McKinney, M. L. 2002b. Influence of settlement time, human population, park shape and age, visitation and roads on the number of alien plant species in protected areas in the USA. *Diversity and Distributions* 8: 311–318.
- Meiners, S. J., M. L. Cadenasso and S. A. Pickett. 2004. Beyond biodiversity: individualistic controls of invasion in a self-assembled community. *Ecology Letters* 7: 121–126.
- Mendelson, J. R., K. R. Lips, R. W. Gagliardo, G. B. Rabb, J. P. Collins, J. E. Diffendorfer, P. Daszak, D. Ibáñez, K. C. Zippel, D. P. Lawson, K. M. Wright, S. N Stuart, C. Gascon, H. R. Silva, P. A. Burrowes, R. L. Joglar, E. La Marca, S. Lötters, L. H. du Preez, C. Weldon, A. Hyatt, J. V. Rodriguez-Mahecha, S. Hunt, H. Robertson, B. Lock, C. J. Raxworthy, D. R. Frost, R. C. Lacy, R. A. Alford, J. A. Campbell, G. Parra-Olea, F. Bolaños, J. J. C. Domingo, T. Halliday, J. B. Murphy, M. H. Wake, L. A. Coloma, S. L. Kuzmin, M. S. Price, K. M. Howell, M. Lau, R. Pethiyagoda, M. Boone, M. J. Lannoo, A. R. Blaustein, A. Dobson, R. A. Griffiths, M. L. Crump, D. B. Wake, and E. D. Brodie Jr. 2006. Biodiversity - Confronting amphibian declines and extinctions. *Science 313*: 48–48.
- Miles, D. B. 1994. Population differentiation in locomotor performance and the potential response of a terrestrial organism to global environmental change. *American Zoologist 34*: 422–436.
- Miller, J. R. and P. Cale. 2000. Behavioural mechanisms and habitat use by birds in a fragmented agricultural landscape. *Ecological Applications 10*: 1732–1748.
- Mitchell, J. F. B., J. Lowe, R. A. Wood, and M. Vellinga. 2006. Extreme events due to human-induced climate change. *Philosophical Transactions of the Royal*

Society a-Mathematical Physical and Engineering Sciences 364: 2117–2133.

- Monello, R. J., J. J. Dennehy, D. L. Murray, and A. J. Wirsing. 2006. Growth and behavioral responses of tadpoles of two native frogs to an exotic competitor, *Rana catesbeiana*. Journal of Herpetology 40: 403–407.
- Murcia, C. 1995. Edge effects in fragmented forests: implications for conservation. *Trends in Ecology and Evolution 10*: 58–62.
- Myhre, K., M. Cabanac and G. Myhre. 1977. Fever and behavioural temperature regulation in the frog *Rana* esculenta. Acta Physiologica Scandinavica 101: 219– 229.
- Naab, F., M. Volcomirsky, A. Burlón, M. E. Caraballo, M. Debray, J. M. Kesque, A. J. Kreiner, M. J. Ozafrán, J. A. Schuff, P. Stoliar, M. E. Vázquez, J. Davidson, M. Davidson, and T. M. F. Schroeder. 2001. Metabolic alterations without metal accumulation in the ovary of adult *Bufo arenarum* females, observed after long-term exposure to zn21, followed by toxicity to embryos. *Archives of Environmental Contamination and Toxicology 41*: 201–207.
- Navas, C. A. 1996a. The effect of temperature on the vocal activity of tropical anurans: a comparison of high and low-elevation species. *Journal of Herpetology 30*: 488–497.
- Navas, C. A., 1996b. Implications of microhabitat selection and patterns of activity on the thermal ecology of high elevation neotropical anurans. *Oecologia* 108: 617–626.
- Navas, C. A., M. M. Antoniazzi, H. Suzuki, and C. Jared. 2007a. Physiological basis for diurnal activity in dispersing juvenile *Bufo granulosus* in the Caatinga, a Brazilian semi-arid environment. *Comparative Biochemistry and Physiology 147A*: 647–657.
- Navas, C. A., J. G. Chaui-Berlinck, J. E. P. W. Bicudo, V. Pivello, and M. Martins. 2007b. Comparative biochemistry and physiology in Brazil: A critical appraisal. *Comparative Biochemestry and Physiology 147A*: 586–593.
- Neckel-Oliveira, S. and C. Gascon. 2006. Abundance, body size and movement patterns of a tropical treefrog in continuous and fragmented forests in the Brazilian Amazon. *Biological Conservation* 128: 308–315.
- Parmesan, C. 2006. Ecological and evolutionary responses to recent climate change. Annual Review of Ecology, Evolution, and Systematics 37: 637–669.
- Parris, M. J., E. Reese and A. Storfer. 2006. Antipredator behavior of chytridiomycosis-infected northern leopard frog (*Rana pipiens*) tadpoles. *Canadian Journal of Zoology* 84: 58–65.
- Pearl, C. A., M. J. Adams, G. S. Schuytema, and A. V. Nebeker. 2003. Behavioral responses of anuran larvae to chemical cues of native and introduced predators

in the Pacific Northwestern United States. *Journal of Herpetology* 37: 572–576.

- Pickford, D. B. and I. D. Morris. 2003. Inhibition of gonadotropin-induced oviposition and ovarian steroidogenesis in the African clawed frog (*Xenopus laevis*) by the pesticide methoxychlor. *Aquatic Toxicology* 62: 179–194.
- Pineda, E. and G. Halffter. 2004. Species diversity and habitat fragmentation: frogs in a tropical montane landscape in Mexico. *Biological Conservation 117*: 499–508.
- Pinto, L. P. and M. C. W. Brito. 2005. Dinâmica da perda da biodiversidade na Mata Atlântica brasileira: uma introdução. Pp. 27-30 in C. Galindo-Leal and I. G. Câmara (eds.), Mata Atlântica: biodiversidade, ameaças e perspectivas. Belo Horizonte, Fundação SOS Mata Atlântica, Centro de Ciências Aplicadas à Biodiversidade.
- Pombal, J. P., Jr. and C. F. B. Haddad. 1999. Frogs of the genus *Paratelmatobius* (Anura: Leptodactylidae) with descriptions of two new species. *Copeia 1999*: 1014– 1026.
- Porter, W. P., S. Budaraju, W. E. Stewart, and N. Ramankutty. 2000. Physiology on a landscape scale: applications in ecological theory and conservation practice. *American Zoologist* 40: 1175–1176.
- Pough, F. H. and T. L. Taigen. 1990. Metabolic correlates of the foraging and social behavior of dart-poison frogs. Animal Behaviour 39: 145–155.
- Pounds, J. A. and M. L. Crump. 1994. Amphibian declines and climate disturbance: The case of the Golden Toad and the Harlequin Frog. *Conservation Biology* 8: 72–85.
- Pounds, J. A., A. C. O. Q. Carnaval, R. Puschendorf, C. F. B. Haddad, and K. L. Masters. 2006b. Responding to amphibian loss. *Science* 314: 1541–1541.
- Pounds, J. A., M. R. Bustamante, L. A. Coloma, J. A. Consuegra, M. P. L. Fogden, P. N. Foster, E. La Marca, K. L. Masters, A. Merino-Viteri, R. Puschendorf, S. R. Ron, G. A. Sánchez-Azofeita, and B. E. Young. 2006a. Widespread amphibian extinctions from epidemic disease driven by global warming. *Nature 439*: 161– 167.
- Pukala, T. L., J. H. Bowie, V. M. Maselli, I. F. Musgrave, and M. J. Tyler. 2006. Host-defense peptides from the glandular secretions of amphibians: structure and activity. *Natural Products Report* 23: 368–393.
- Rank, N. E. and E. P. Dahlhoff. 2002. Allele frequency shifts in response to climate change and physiological consequences of allozyme variation in a montane insect. *Evolution* 56: 2278–2289.
- Reading, C. J. 2007. Linking global warming to amphibian declines through its effects on female body condition and survivorship. *Oecologia* 151: 125–131.

- Relyea, R. A. and N. Millis. 2001. Predator induced stress makes the pesticide carbaryl more deadly to gray tree frogs tadpoles (*Hyla versicolor*). Proceedings of the National Academy of Sciences 98: 2491–2496.
- Retallick, R. W. R., H. McCallum and R. Speare. 2004. Endemic infection of the amphibian chytrid fungus in a frog community post-decline. *Plos Biology* 2: 1965– 1971.
- Rocha-Miranda, F., M. J. Martins Silva and A. F. Mendonça. 2006. First occurrence of bullfrogs (*Rana catesbeiana*) in Federal District, Central Brazil. *Froglog* 74: 2–3.
- Rollins-Smith, L. A. and J. M. Conlon. 2005. Antimicrobial peptide defenses against chytridiomycosis, an emerging infectious disease of amphibian populations. *Developmental and Comparative Immunology 29*: 589–598.
- Rollins-Smith, L. A., L. K. Reinert, C. J. O'Leary, L. E. Houston, and D. C. Woodhams. 2005. Antimicrobial peptide defenses in amphibian skin. *Integrative and Comparative Biology* 45: 137–142.
- Rollins-Smith, L. A., C. Carey, J. Longcore, J. K. Doersam, A. Boutte, J. E. Bruzgal, and J. M. Conlon. 2002a. Activity of antimicrobial skin peptides from ranid frogs against *Batrachochytrium dendrobatidis*, the chytrid fungus associated with global amphibian declines. *Developmental and Comparative Immunology* 26: 471–479.
- Rollins-Smith, L. A., J. K. Doersam, J. E. Longcore, S. K. Taylor, J. C. Shamblin, C. Carey, and M. A. Zasloff. 2002b. Antimicrobial peptide defenses against *Batrachochytrium dendrobatidis*, the chytrid fungus associated with global amphibian declines. *Faseb Journal* 16: A291–A291.
- Ron, S. R. 2005. Predicting the distribution of the amphibian pathogen *Batrachocytrium dendrobatidis* in the New World. *Biotropica* 37: 209–221.
- Rowe, C. L., W. A. Hopkins and V. R. Coffman. 2001. Failed recruitment of southern toads (*Bufo terrestris*) in a trace element-contaminated breeding habitat: direct and indirect effects that may lead to a local population sink. Archives of Environmental Contamination and Toxicology 40: 399–405.
- Rowe, C. L., W. J. Sadinski and W. A. Dunson. 1992. Effects of acute and chronic acidification on three larval amphibians that breed in temporary ponds. *Archives of Environmental Contamination and Toxicology 23*: 339–350.
- Rowe, C. L., O. M. Kinney, A. P. Fiori, and J. D. Congdon. 1996. Oral deformities in tadpoles (*Rana catesbeiana*) associated with coal ash deposition: effects on grazing ability and growth. *Freshwater Biology* 36: 723–730.
- Rowe, C. L., O. M. Kinney, R. D. Nagle, and J. D. Congdon. 1998. Elevated maintenance costs in an

anuran (*Rana catesbeiana*) exposed to a mixture of trace elements during embryonic and early larval periods. *Physiological Zoology* 35: 218–223.

- Sadinski, W. J. and W. A. Dunson. 1992. A multilevel study of low pH on amphibians of temporary ponds. *Journal* of Herpetology 26: 413–422.
- Saunders, D. A., R. J. Hobbs and C. R. Margules. 1991. Biological consequences of ecosystem fragmentation: a review. *Conservation Biology* 5: 18–32.
- Sax, D. and S. D. Gaines. 2003. Species diversity: from global decreases to local increases. *Trends in Ecology* and Evolution 18: 561–566.
- Schiesari, L., B. Grillitsch and H. Grillitsch. 2007. Biogeographic biases in research and their consequences for linking amphibian declines to pollution. *Conservation Biology* 21: 465–471.
- Sears, M. W. and M. J. Angilletta. 2004. Body size clines in *Sceloporus* lizards: Proximate mechanisms and demographic constraints. *Integrative and Comparative Biology* 44: 433–442.
- Sherman, E., L. Baldwin, G. Fernandez, and E. Deurell. 1991. Fever and thermal tolerance in the toad *Bufo* marinus. Journal of Thermal Biology 16: 297–302.
- Silvano, D. L. and M. V. Segalla. 2005. Conservation of Brazilian amphibians. *Conservation Biology* 19: 653–658.
- Simmaco, M., A. Boman, M. L. Mangoni, G. Mignogna, R. Miele, D. Barra, and H. G. Boman. 1997. Effect of glucocorticoids on the synthesis of antimicrobial peptides in amphibian skin. *Federation of European Biochemical Societies Letters* 416: 273–275.
- Simon, M. P., I. Vatnick, H. A. Hopey, K. Butler, C. Korver, C. Hilton, R. S. Weimann, and M. A. Brodkin. 2002. Effects of acid exposure on natural resistance and mortality of adult *Rana pipiens*. *Journal of Herpetology* 36: 697–699.
- Skelly, D. K. 1992. Field evidence for a cost of behavioral antipredator response in a larval amphibian. *Ecology* 73: 704–708.
- Smith, M. A. and D. M. Green. 2005. Dispersal and the metapopulation paradigm in amphibian ecology and conservation: are all amphibian populations metapopulations? *Ecography* 28: 110–128.
- SOS Mata Atlântica. 1998. Atlas da evolução dos remanescentes florestais e ecossistemas associados no domínio da Mata Atlântica no período 1990-1995. Fundação SOS Mata Atlântica, São Paulo, Brazil.
- Spotila, J. R., M. P. O'Connor and G. S. Bakken. 1992.
 Biophysics of heat and mass transfer. Pp. 59–80 in M.
 E. Feder and W. W. Burggren (eds.), *Environmental Physiology of the Amphibians*. Chicago, The University of Chicago Press.

- Stevenson, R. D., S. R. Tuberty, P. L. Defur, and J. C. Wingfield. 2005. Eco-physiology and conservation: The contribution of endocrinology and immunology. Introduction to the symposium. *Integrative and Comparative Biology* 45: 1–3.
- Stillman, J. H. 2003. Acclimation capacity underlies susceptibility to climate change. Science 301: 65–65.
- Stohlgren, T. J., D. Binkley, G. W. Chong, M. A. Kalkhan, L. D. Schell, K. A. Bull, Y. Otsuki, G. Newman, M. Bashkin, and Y. Son. 1999. Exotic plant species invade hotspots of native plant diversity. *Ecological Mono*graphs 69: 25–46.
- Sutherst, R. W., R. B. Floyd and G. F. Maywald. 1996. The potential geographical distribution of the cane toad, *Bufo marinus* L in Australia. *Conservation Biology 10*: 294–299.
- Taigen, T. L. and F. H. Pough. 1985. Metabolic correlates of anuran behaviour. American Zoologist 25: 987–997.
- Taigen, T. L., S. B. Emerson and F. H. Pough. 1982. Ecological correlates of anuran exercise physiology. *Oecologia* 52: 49–56.
- Toledo, L. F., C. F. B. Haddad, A. C. O. Q. Carnaval, and F. B. Britto. 2006a. A Brazilian anuran (Hylodes magalhaesi: Leptodactylidae) infected by Batrachochytrium dendrobatidis: a conservation concern. Amphibian and Reptile Conservation 4: 17–21.
- Toledo, L. F., F. B. Britto, O. G. S. Araújo, L. M. O. Giasson, and C. F. B. Haddad. 2006b. The occurrence of *Batrachochytrium dendrobatidis* in Brazil and the inclusion of 17 new cases of infection. *South American Journal of Herpetology 1*: 185–191.
- Tryjanowski, P., T. Sparks, M. Rybacki, and L. Berger. 2006. Is body size of the water frog *Rana esculenta* complex responding to climate change? *Naturwissenschaften* 93: 110–113.
- Turner, I. M. 1996. Species loss in fragments of tropical rain forest: a review of the evidence. *Journal of Applied Ecology 33*: 200–209.
- Vallan, D. 2000. Influence of forest fragmentation on amphibian diversity in the nature reserve of Ambohitantely, highland Madagascar. *Biological Conservation* 96: 31-43.
- Vanzolini, P.E. and E. E. Williams. 1970. South American anoles; the geographic differentiation and evolution of the Anolis chrysolepis species group (Sauria, Iguanidae). Arquivos de Zoologia, São Paulo 19: 1–240.
- Weygoldt, P. 1989. Changes in the composition of mountain stream frog communities in the Atlantic mountains of Brazil: frogs as indicators of environmental deteriorations? *Studies on Neotropical Fauna and Environment 243*: 249–255.
- Wikelski, M. and S. J. Cooke. 2006. Conservation physiology. Trends in Ecology and Evolution 21: 38–46.

- Williamson, M.1996. *Biological Invasions*. London, Chapman and Hall.
- Woinarski, J. C. Z., J. C. McCosker, G. Gordon, B. Lawrie, C. James, J. Augusteyn, L. Slater, and T. Danvers. 2006. Monitoring change in the vertebrate fauna of central Queensland, Australia, over a period of broadscale vegetation clearance, 1973–2002. Wildlife Research 33: 263–274.
- Wolf, B. 2000. Global warming and avian occupancy of hot deserts; a physiological and behavioral perspective. *Revista Chilena de Historia Natural* 73: 395–400.
- Woodhams, D. C. and R. A. Alford. 2005. Ecology of chytridiomycosis in rainforest stream frog assemblages of tropical Queensland. *Conservation Biology* 19: 1449–1459.
- Woodhams, D. C., R. A. Alford and G. Marantelli. 2003. Emerging disease of amphibians cured by elevated body temperature. *Diseases of Aquatic Organisms 55:* 65–67.
- Woodhams, D. C., L. A. Rollins-Smith, C. Carey, L. Reinert, M. J. Tyler, and R. Alford. 2006a. Population

trends associated with skin peptide defenses against chytridiomycosis in Australian frogs. *Oecologia 146*: 531–540.

- Woodhams, D. C., J. Voyles, K. R. Lips, C. Carey, L. A. Rollins-Smith, and R. Alford. 2006b. Predicted disease susceptibility in a Panamanian amphibian assemblage based on skin peptide defenses. *Journal of Wildlife Disease* 42: 207–218.
- Young, B. E., K. R. Lips, J. K. Reaser, R. Ibanez, A. W. Salas, J. R. Cadeno, L. A. Coloma, S. Ron, E. La Marca, J. R. Meyer, A. Munos, F. Bolanos, G. Chaves, and D. Romo. 2001. Population declines and priorities for amphibian conservation in Latin America. *Conservation Biology* 15: 1213–1223.
- Zimmerman, B.L. and R. O. Bierregaard. 1986. Relevance of the equilibrium-theory of island biogeography and species area relations to conservation with a case from Amazonia. *Journal of Biogeography* 13: 133–143.
- Zuidema, P.A., J. A. Sayer and W. Dijkman. 1996. Forest fragmentation and biodiversity: the case for intermediate-sized conservation areas. *Environmental Conservation* 23: 290–297.